

Popular Article

Paratuberculosis in cattle

Gaurav agrawal^{1*}, Dr.Nazeer Mohammad², Dilip Singh Meena³

DOI: <https://doi.org/10.5281/zenodo.6659666>

Introduction

In India, paratuberculosis, also known as Johne's disease (JD), is endemic in domestic livestock and was first reported in Hisar, India, in 1913. National estimates of the prevalence of MAP in India are still unavailable (Kumar *et al.*, 2007, Sharma *et al.*, 2008). John's disease causes enormous economic losses and has a significant impact on the livestock industry due to premature culling of animals, reduced weight gain, reduced feed efficiency, and high morbidity (Kaur *et al.*, 2011); reduced carcass value, reduced milk production, increased susceptibility to mastitis and reproductive disorders, resulting in increased calving intervals, decreased fertility, and additional veterinary costs (Hasonova & Pavlik, 2006). Paratuberculosis causes significant economic losses in the dairy industry in the United States, estimated at more than \$200 million per year (Groenendaal *et al.*, 2015), and thus has a significant impact in both developing and developed countries. No country claims to be free on MAP (Yue *et al.*, 2016). Paratuberculosis, often known as Johne's disease, a chronic intestinal disease that mostly affects ruminants and camelids. In animals, paratuberculosis causes intermittent or chronic diarrhoea, chronic weight loss, and generalized stinginess (Whitlock and Buergelt, 1996).

Etiology

The disease is caused by *Mycobacterium avium* subspecies paratuberculosis (MAP). The organism is acid-fast, aerobic, and slow-growing, and its in-vitro growth requires exogenous mycobactin (Merkal and Curran 1974, Chiodini *et al.* 1984).

Epidemiology

The disease occurs in most parts of the world and the prevalence seems to be increasing in some countries. Paratuberculosis is predominant in cattle and sheep in temperate climates with adequate rainfall and ground waters and in some humid, tropical areas. The incidence of paratuberculosis is high in animals kept intensively under environmental and husbandry conditions which are conducive to the spread of the infection (Chiodini *et al.*, 1984a).

Post Graduate Institute of Veterinary Education and Research (PGIVER), Jaipur, RAJUVAS, Bikaner, Rajasthan

Mode of transmission

The important route of MAP transmission is the fecal-oral route between infectious cows and susceptible young calves via ingestion of contaminated milk, water, or uptake from the environment (Lombard 2011). Recent studies on the risk of faecal shedding in calves born to faecal culture positive dams or fed MAP-containing colostrum failed to confirm these proposed risk factors (Pithua *et al* 2012). In cows in the subclinical and clinical stages of the infection, intra-uterine transmission of MAP has been described (McQueen and Russell 1979, Sweeney *et al* 1996). The respiratory tract of cattle was proposed as an additional route of infection for MAP (Eisenberg *et al* 2012).

Clinical signs

In cattle the disease is characterized by chronic and intermittent diarrhoea that is not responsive to treatment, oedema of the throat and abdomen, loss of coat colour, emaciation and eventual death. Due to the chronic nature of the disease, clinical manifestations of paratuberculosis can appear as late as 3 to 5 years after infection (Riemann and Abbas, 1983; Chiodini *et al.*, 1984a).

Post mortem findings.

The ileum often has a severely thickened and corrugated appearance due to the granulomatous infiltrate. Histologically, *M. paratuberculosis* is found in macrophages which infiltrate into the lamina propria of the intestine (Kubo *et al.*, 1983). In cattle, there is no caseation, calcification, or fibrosis associated with paratuberculosis lesions (Hines *et al.*, 1995).

Diagnosis.

Primary diagnosis can be made based on clinical signs, postmortem lesion, and histopathology, and it can be confirmed by a battery of diagnostic tests such as faecal culture, acid-fast staining, agar gel immunodiffusion (AGID), complement fixation test (CFT), delayed type hypersensitivity (DTH), interferon gamma assay, and enzyme linked immune-sorbent assay (ELISA). Culture-based methods are the "gold standard" for MAP infection diagnosis. These methods, however, are time-consuming and have a low sensitivity. Recently, molecular biological methods such as polymerase chain reaction (PCR) have been developed to detect MAP infection in animals (Fang *et al.*, 2002).

Prevention and control

The prevention and management of John's disease in humans and animals, particularly dairy animals, is critical, and requires a robust control mechanism to prevent disease spread within species and between people and animals. One of the most limiting issues is the lack of a reliable, cost-effective, and quick diagnostic test (s).

Vaccination

Vaccination of cattle against paratuberculosis is not routinely recommended. Although it may have served a purpose in the past, recent information indicates it is of limited value in controlling *M. paratuberculosis* infections, causes a false sense of security in owners, is a serious health risk for veterinarians and prevents use of serologic tests in a herd (Collins, 1994).

Treatment

Currently, no antimicrobials are approved for the treatment of Johne's disease. *M. paratuberculosis* is more resistant to chemotherapeutic agents in vitro than *M. tuberculosis* so that prospects for suitable treatment are poor. Because of this lack of efficacy and the failure of any of the antimicrobials to provide a bacteriological cure, treatment is not recommended. (Radostits and Blood, 1994)

References

- Merkal R S and Curran B. (1974). Growth and metabolic characteristics of *Mycobacterium paratuberculosis*. *Applied Microbiology* 28
- Whitlock R H and Buergelt C. (1996). Preclinical and clinical manifestations of paratuberculosis (including pathology). *Veterinary Clinics of North America: Food Animal Practice* 12(2)
- Eisenberg S W F, Nielen M, Santema W, Houwers D J, Heederik D and Koets A P. (2010). Detection of spatial and temporal spread of *Mycobacterium avium subsp. paratuberculosis* in the environment of a cattle farm through bio-aerosols. *Veterinary microbiology* 143(2-4): 284-92.
- Lombard JE. (2011). Epidemiology and economics of paratuberculosis. *Veterinary Clinics of North America: Food Animal Practice* 27(3): 525-35.
- Pithua P and Kollias N S. (2012). Estimated prevalence of caprine paratuberculosis in boer goat herds in Missouri. *USA Veterinary Medicine International* 2012: 674085.
- McQueen D S and Russell E G. (1979). Culture of *Mycobacterium paratuberculosis* from bovine foetuses. *Australian Veterinary Journal* 55(4): 203-04.
- Sweeney R W. (1996). Transmission of paratuberculosis. *Veterinary Clinics of North America: Food Animal Practice* 12: 305-12.
- Fang, Y., Wu, W.-H., Pepper, J.L., Larsen, J.L., Marras, S.A., Nelson, E.A., Epperson, W.B. and Christopher-Hennings, J. (2002). Comparison of real-time, quantitative PCR with molecular beacons to nested PCR and culture methods for detection of *Mycobacterium avium subsp. paratuberculosis* in bovine fecal samples. *J. Clin. Microbiol* 40: 287-291.
- Hasonova, L. & Pavlik, I. (2006). Economic impact of paratuberculosis in dairy cattle herds: a review. *Veterinarni Medicina*, 51(5), 193 - 211.

- Kaur, P., Folia, G., Singh, S.V., Patil, P.K., Ravi Kumar, G.V.P.P.S. & K.S. Sandhu (2011). Molecular epidemiology of *Mycobacterium avium* subspecies *paratuberculosis*: IS900 PCR identification and IS1311 polymorphism analysis from ruminants in the Punjab region of India. *Comparative Immunology, Microbiology and Infectious Diseases*, 34, 163 - 169.
- Yue, R., Liu, C., Barrow, P., Liu, F., Cui, Y., Yang, L., Zhao, D., & Zhou, X. (2016). The isolation and molecular characterization of *Mycobacterium avium* subsp. *paratuberculosis* in Shandong province, China. *Gut Pathogens*, 8(9), 1-9.
- Groenendaal, H., Zagmutt, F. J., Patton, E. A., & Wells, S. J. (2015). Cost-benefit analysis of vaccination against *Mycobacterium avium* ssp. *paratuberculosis* in dairy cattle, given its cross-reactivity with tuberculosis tests. *Journal of Dairy Science*, 98, (9) 6070 - 6084.
- Riemann H.P., Abbas B. (1983): Diagnosis and control of bovine paratuberculosis. *Adv. Vet. Sci. Com. Med.*, 27, 481–506.
- Chiodini R.J., Van Kruiningen H.J., Merkal R.S. (1984a): Ruminant paratuberculosis (Johne's disease): the current status and future prospects. *Cornell Vet.*, 74, 218–262.
- Kumar, P., Singh, S.V., Bhatiya, A.K. Sevilla, L., Singh, A.V., Whittington, R.A., Juste, R.A., Gupta, V.K., Singh, P., und Sohal, J.S. and Vihan, V.S (2007). Juvenile Capri Paratuberculosis (JCP) in India Incidence and characterization by six diagnostic tests. *Small Rumin. Res.*, 73: 45-53.
- Sharma, G. Singh, S.V., Sevilla, I. Singh, A.V., Whittington, R.J., Juste, R.A., Kumar, S., Gupta, V.K., Singh, P, Sohal, J.S. and Vihan, V.S. (2008). Evaluation of indigenous milk ELISA with m-culture and m-PCR for the diagnosis of bovine Johne's disease (BID) in lactating Indian dairy cattle. *Res Vet. Sci.*, 84: 30-37.
- Hines M.E., Kreeger J.M., Herron A.J. (1995): Mycobacterial infections of animals: pathology and pathogenesis. *Lab. Anim. Sci.*, 45, 334–351.
- Kubo M., Moriwaki M., Watase H. (1983): Electron microscopic observations on the intestine of a cow with Johne's disease. *Jpn. J. Nat. Sci.*, 45, 259–262.
- Collins M.T. (1994): Clinical approach to control of bovine paratuberculosis. *J. Am. Vet. Med. Assoc.*, 204, 208–210.
- Radostits, O.M., Blood, D.C. and Gay, C.C. (1994) *Veterinary Medicine, a Text Book of the Disease of Cattle, Sheep, Goats, Pigs and Horses*. 8th Edition, Bailliere, Tindall, London, 1015-1026.

Cite as

Gaurav agrawal^{1*}, Dr.Nazeer Mohammad², Dilip Singh Meena³. (2022). Paratuberculosis in cattle. *The Science World a Monthly E Magazine*, 2(6), 675–678.
<https://doi.org/10.5281/zenodo.6659666>

